PROTEABIOSCIENCES

GPR™-800

ADVANCED PROTEIN RECOVERY FROM GEL SAMPLES

USERS MANUAL

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Use of the GPR-800 involves the use of and potential exposure to hazardous materials, and therefore, the system is intended for use only by professionals with adequate training and experience in handling of the dangerous materials involved. Protea cannot be held responsible for any injuries to persons or property or for any other damage resulting from the improper use of this equipment or its use in any manner not specifically described in this manual. Furthermore, Protea disclaims all responsibility and liability in connection with any accidents arising from any use of the GPR-800 in situations where inadequate safety procedures are in place or where such procedures have not been followed.

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WARRANTY: Products are Warranted to be free of defects in materials and workmanship, and to perform to the published specifications, for a period of one year from the date of the product's shipment (see current Protea Biosciences, Inc. catalog for additional warranty language and Terms of Sale).

Preface

This user's manual is written for use with the Protea Biosciences, Inc. (Protea) Gel Protein Recovery System 800 (hereafter "GPR-800"). This device's intended use is to serve as a platform for developing the gel protein recovery process. The audience of this device is the intended eventual end users (i.e., practitioners) of the device.

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1 Introduction

This chapter contains introductory information on the Gel Protein Recovery System (GPR-800), including specifications. It also provides information about this User's Manual.

1.1 INTRODUCING THE GPR-800

The GPR-800 is intended for use in a research laboratory to support extraction of intact proteins, peptide mixtures from protein digests, and other biomolecules from polyacrylamide gel plugs by electroelution or electroextraction.*

The GPR-800 is designed to efficiently recover biomolecules from acrylamide gel plugs via a decoupled sample recovery process that minimizes set-up, operation time, and user interaction.

The operator begins the process by excising poylacrylamide gel plugs and placing them into the designated receptacles. The GPR-800 is primed by the addition of GPR Electroelution Buffer into the electrode reservoirs. After completing the system setup, the operator starts the sample recovery process using the front panel interface on the GPR-800. After electroelution the recovered protein samples are transferred to vials for subsequent desalting and detergent degradation using reversed phase functionalized pipette tips (e.g. LithTips).

For instructions on general use of the GPR-800, see the General Operation section starting on page 23 of this manual.

1.2 ABOUT THE USER'S MANUAL

This manual contains information on the features and functions of the GPR-800. It provides instructions on operating the unit. Read the Safety Information starting on page 11 prior to setting up or operating the GPR-800, as this section contains information necessary for the safe operation of the unit. Important information contained in this manual is marked as follows:

WARNING: Impending Danger!
May cause death or physical injury.

CAUTION: Dangerous Situation!
May cause equipment/material damage or data loss.

NOTE: Helpful additional information and tips for use.

*patent pending

1.3 TECHNICAL SPECIFICATIONS

The GPR-800 has the specifications listed below.

1.3.1 Physical Dimensions*

Physical	U.S. Units	SI Units
Dimensions (W x H x D)	13 in x 9 in x 14 in	33 cm x 23 cm x 36 cm
Weight	20 lb	9.0 kg

* Dimensions and weight are approximate.

1.3.2 Environmental Ranges

Environmental	Operation	Storage
Temperature**	10 to 50°C (50 to 122°F)	10 to 50°C (50 to 122°F)
Relative Humidity	35 to 85% non-condensing	35 to 85% non-condensing

**Exposing the system to direct sunlight is not recommended.

1.3.3 Consumables Storage Specifications

CAUTION: Consumable materials to be used with the GPR-800 must be stored in accordance with the requirements set forth on their labels and packaging.

1.3.4 Electrical Specifications

Input Voltage	100-240 V
Current	0.30A maximum
Frequency	50/60 Hz
Fuses	Fast-acting, 250 V, 5x20 mm Mains Fusing: 1A (Bell Fuse P/N 5SF 1-R, qty 2) High Voltage DC Supply Fusing: 200 mA (Bell Fuse P/N 5SF 200-R, qty 2)

CAUTION: Fuses are not serviceable by the end user. Please call Protea Technical Support.

1.3.5 Chemical Compatibility

Exterior surfaces of the device enclosure are ABS, acetal, aluminum, EPDM, neoprene, polycarbonate, and polyester. Care should be taken to ensure chemicals used in operating, cleaning, and maintaining the system do not remain in contact with materials with which they are incompatible, as system damage may occur.

1.3.6 Flammable Liquids

Flammable liquids may be used during operation of the GPR-800. Follow your institutional requirements for the handling and disposal of the flammable or otherwise hazardous liquids.

CAUTION: The total volume of flammable liquids used in the GPR-800 per chip run should not exceed 1 mL during proper operation of the device.

2 Safety

This chapter contains safety information necessary for the safe operation of the GPR-800. Read and understand this chapter prior to setting up or operating the unit.

Always adhere to the safety standards regulating your operating environment. Also adhere to the safety rules outlined below in addition to your specific standards and guidelines.

WARNING: Use of the system involves the potential use of and exposure to potentially harmful materials. Only use the system if you have adequate training and experience in handling such materials.

WARNING: Turn off electrical power to and unplug the system prior to any factory or end-user maintenance or cleaning.

WARNING: Only operate the system when the system is fully assembled.

WARNING: Do not operate the system if unit safety lid and electrode hinge are not properly installed, or if the unit safety lid or electrode hinge are damaged or broken.

WARNING: Do not attempt to disassemble, alter, override, or otherwise defeat safety switches or features of the system. Doing so may result in injury or death.

WARNING: Use caution when coming in close contact with the electrodes and other components within the system, as these components could cause physical harm, especially in cases of a malfunction.

WARNING: Use caution when working near the operating system, as the system contains high voltages.

WARNING: Remove power from the system immediately upon fluid spills on or near the system. Do not operate the system after fluid spills have occurred.

WARNING: No modification of this equipment is permitted.

WARNING: Do not operate the system if damage or abnormal/erratic operation has been detected.

CAUTION: Keep open containers of fluid away from the system.

CAUTION: Keep areas around the system clean at all times.

CAUTION: Maintain at least 6 inches of unobstructed space around the unit to ensure proper use of the system.

2.1 SAFETY STOPS

The GPR-800 employs automatic safety stops as outlined below. When an automatic safety stop is required the system enters an abort mode where all voltage is removed from the electrodes, or the user is prevented from applying voltage to the electrodes.

2.1.1 Unit Safety Cover

A safety cover is provided to protect the user from coming into contact with the high voltage elements of the GPR-800. The system is designed to automatically remove voltage from all electrodes when the cover is opened or removed.

Figure 2-1. GPR-800 with Safety Cover closed

Figure 2-2. GPR-800 with Safety Cover open

WARNING: Do not attempt to run the system without the safety cover closed, as doing so may cause physical injury.

NOTE: Electroelution voltages will not be applied to reservoir samples if the safety cover is not closed. If the channel LEDs are blinking, the safety cover may be open.

2.1.2 Chip Holder

The chip holder protects the user from touching the high voltage electrodes inside the GPR-800 when it is in place and the safety cover is closed. The system is designed to automatically remove voltage from all electrodes when the chip holder is removed from or improperly installed in the system. The chip holder contains a magnetic safety lock that must be engaged by properly placing the chip holder into position for the GPR-800 to operate under high voltage.

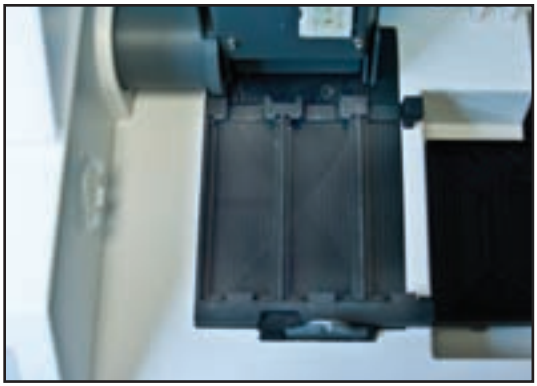

Figure 2-3. GPR-800 with Chip Holder installed

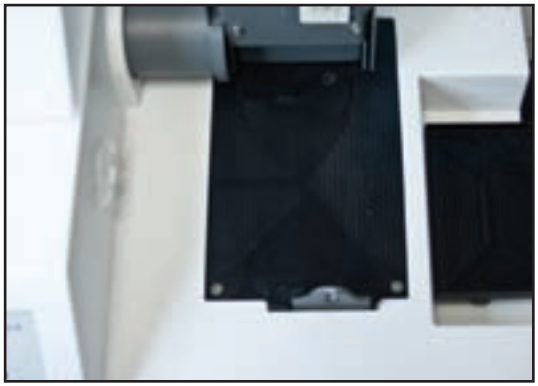

Figure 2-4. GPR-800 with Chip Holder removed

WARNING: Do not attempt to run the system without the chip holder in place, as doing so may cause physical injury.

NOTE: Electroelution voltages will not be applied to reservoir samples if the chip holder is not installed on the chip platform. If the channel LEDs are blinking, the chip holder may not be installed.

2.2 ELECTRICAL SAFETY

WARNING: Connect the system to an electrical power source with earth ground using the three-prong, grounded power cord. Ensure that the supply outlet is grounded and that the voltage of your main power supply matches the voltage of the unit.

WARNING: Turn off electrical power to the system and unplug the power supply cord before performing any cleaning, service, or maintenance on the GPR-800.

WARNING: Remove power from the system immediately upon fluid spills on or near the system. Do not operate the unit if fluid spills have occurred.

3 Component Locations

This chapter provides the locations and descriptions of the components that make up the GPR-800.

3.1 EXTERNAL AND INTERNAL PARTS

Figure 3-1. Shows the external parts located on the front of the GPR-800.

1	**Channel LEDs**	Indicate if the corresponding channels are powered and if they are receiving over 50 µA +/- 5 µA. LED states: • Off - No electroelution voltage provided to channel. • Green - Electroelution voltage provided. Channel is drawing at least 50 µA of current. • Red - Electroelution voltage provided. Channel is drawing less than 50 µA of current. • Blinking - Electroelution process is PAUSED by user or because safety mechanisms are not in place (see section 2.1). No electroelution voltage provided to channel.
2	**Power Status LED**	Indicates whether or not the device is powered.
3	**Timer Relay (Left)**	Controls the time voltage is applied for electrode channels 1-4.
4	**Timer Relay (Right)**	Controls the time voltage is applied for electrode channels 5-8.
5	**Keypad**	Controls the timer relays by starting, pausing, or stopping the process and by allowing the electroelution time setpoint to be modified.

Figure 3-2. Shows the external and internal parts located on GPR-800.

6	**Vial Lift Knob**	Raises and lowers the vial holder for positioning of sample vials.
7	**Vial Lift**	Positions the vial holder and vials below the outlet channel bridges.
8	**Safety Cover**	Protects user from high voltages.
9	**EH (Electrode Hinge)**	Controls the positioning of the electrodes into either the OPEN, RESTING, or SEALED position.
10	**Electrodes**	Provide electroelution voltage (16 electrodes supply 8 channels).
11	**Chip Holder**	See Section 5.
12	**Vial Holder**	See Section 5.
13	**EH Knob**	Interfaces with the EH Catch Plate to position the EH in either the RESTING or SEALED position.
14	**EH Catcher Plate**	Interfaces with the EH Knob to position the EH in either the RESTING or SEALED position.
15	**Leveling Mounts**	Allow the GPR-800 to sit level on a work surface. These mounts can be rotated to adjust the level of the GPR-800.

Figure 3-3. Shows the external parts located on the rear of the GPR-800.

1	**Power Entry Module**	AC power entry port and hard power switch (controls all power to the system). Requires an input from an electrical power source with earth ground using the supplied three-prong, grounded power cord.
2	**USB Connector**	Communication link for the GPR-800 software.

3.2 SYSTEM ACCESSORIES

These accessories organize the GPR-800 consumables and allow them to be loaded onto the unit. The consumables needed for the system during normal use are described in Section 8.

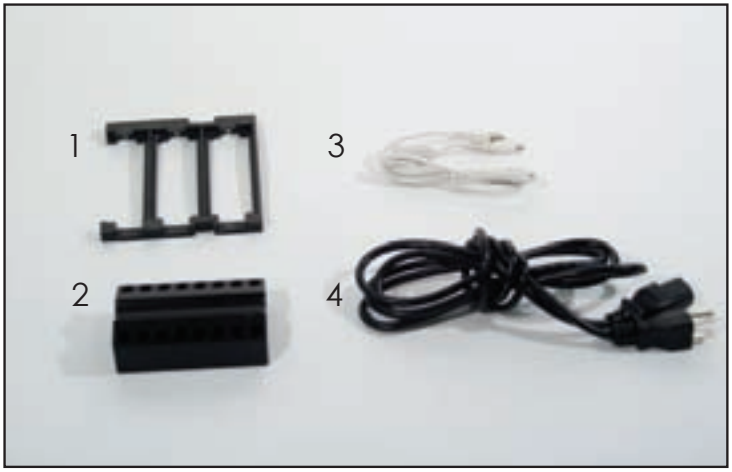

Figure 3-4. Shows the GPR-800 accessories for loading the consumables and for powering and communicating with the GPR-800.

1	**Chip Holder**	Holds the Protein GPRchip.
2	**Vial Rack**	Holds the sample and waste collection vials.
3	**USB Cable**	Connects the GPR-800 to a computer (computer not provided).
4	**Power Cable**	Connects the GPR-800 to an appropriate AC power source.

4 Installation and Preparation

4.1 ACCESSORY EQUIPMENT

The following equipment is not supplied with the GPR-800 but is recommended for use with the system:

- Computer — A computer running the GPR Works software shows the current for each channel in real time. This data can be logged and saved for record keeping and data evaluation purposes. The computer/software feature is not required for operation of the GPR-800.

An appropriate computer will meet the following requirements (at a minimum):

- Windows 2000 / XP or later version
- 64 MB RAM (for Pentium 200MHz processor or equivalent);
  256 MB RAM (for Pentium III/Celeron 600 MHz processor or equivalent),
- 800x600 pixels (for Pentium 200 MHz processor or equivalent);
  1024x768 pixels (for Pentium III/Celeron 600 MHz processor or equivalent), and
- USB interfaceport

4.2 INITIAL SETUP

This chapter provides information on installing and preparing the GPR-800 for initial use. Follow these steps prior to using the GPR-800.

WARNING: Read the Safety information starting on page 11 prior to setting up or operating the GPR-800.

4.2.1 Unpacking and Setup of the GPR-800

Step 1 Unpack System

Unpack the system from its shipping container and place the GPR-800 on a sturdy, level, flat surface. Ensure that the AC power disconnect is accessible when the system is positioned for use.

WARNING: Do not position the system in a location where liquids can spill onto electrical components.

NOTE: Be sure to place the GPR-800 unit on a flat, level, stable surface that can easily support 30 pounds. Adjust the leveling mounts (GPR-800 feet) so that the instrument is secure and level in position.

Step 2 Install AC Power Cord

Ensure that the main power switch on the rear of the unit is in the OFF position.
Unpack the AC power cord from the shipping container, and connect the GPR-800 via the power entry module to an electrical power source with earth ground using the supplied three-prong, grounded power cord. Ensure that the supply outlet is grounded and that the voltage of your main power supply matches the voltage of the unit.

CAUTION: Make sure the outlet is grounded, and that the voltage of your main power supply matches the voltage of the unit.

Step 3 Install GPR Works Software

If using the GPR Works software, install the software on an appropriate computer. Install the software by placing the installation CD into the computer's CD-ROM drive. Software installation should start automatically.

When the installation parameters appear on the computer screen (Figure 4-1), accept the default values. The installation will proceed automatically and may take several minutes. When the installation is complete, press the "Finish" button. If prompted, reboot the computer to complete the installation.

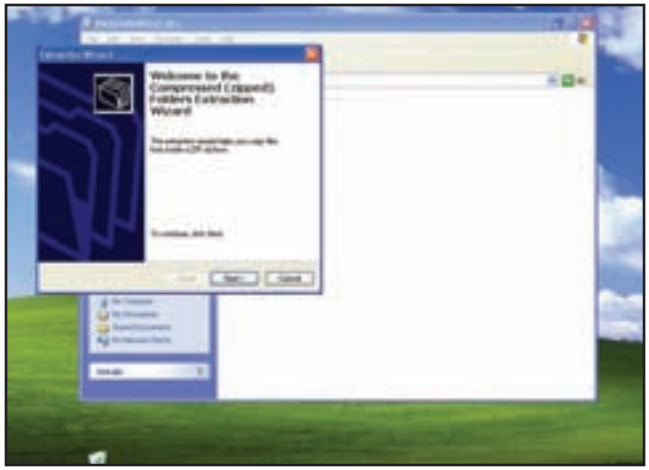

Figure 4-1.
GPR Software Installation Parameters

Step 4 Unpack USB Cable

If using the GPR Works software, unpack the USB cable, and connect the end of the cable with the mini-USB connector to the rear of the GPR-800. Connect the other end of the cable to the computer running the GPR Works software.

Step 5 Enter Calibration Values

Start the GPR Works software on the controlling computer. Click the calibration button. Enter the 8 calibration values for the device being used. Select OK to save the calibration values and dismiss the dialog.

> NOTE: Calibration values are specific to each GPR-800 and are supplied by Protea. The calibration values are located on the back of each GPR-800 on a label. If using more than one GPR-800 with a given computer, the calibration values will need to be appropriately set each time a different GPR-800 is running.

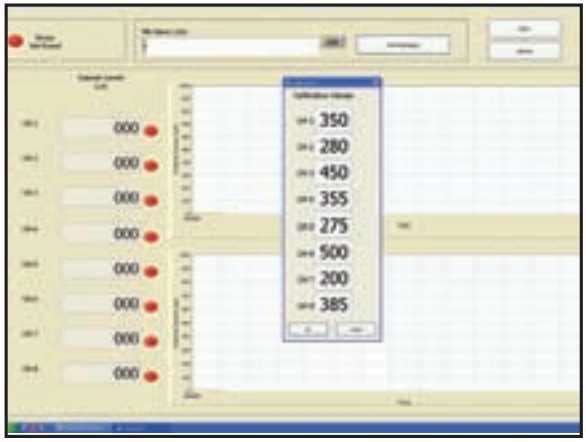

Figure 4-2.
GPR-800 Calibration Value Entry

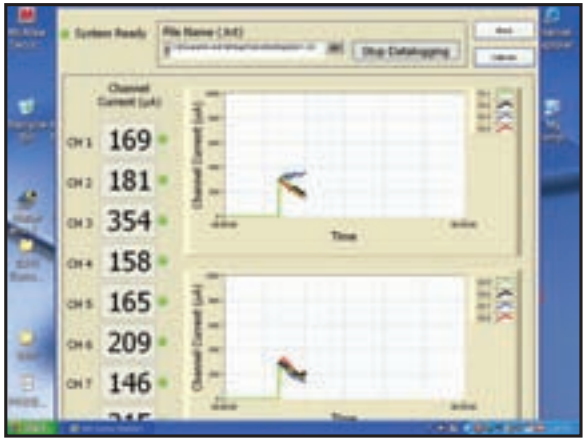

Figure 4-3.
Software Process Screen

Step 6 Power Unit ON

Toggle the main power switch on the rear of the unit. After the unit is powered ON, the GPR-800 Power Status LED should illuminate and the timer relays should turn on.

5 Instructions for Use

This chapter provides an overview of operation of the GPR-800. See the Installation and Preparation instructions in Section 4 for guidance on installing and preparing the GPR-800 for initial use.

Safety Warning and Limitation of Liability

Use of the GPR-800 involves the use of and potential exposure to hazardous materials, and therefore, the system is intended for use only by professionals with adequate training and experience in handling of the dangerous materials involved. Protea cannot be held responsible for any injuries to persons or property or for any other damage resulting from the improper use of this equipment or its use in any manner not specifically described in this manual. Furthermore, Protea disclaims all responsibility and liability in connection with any accidents arising from any use of the GPR-800 in situations where inadequate safety procedures are in place or where such procedures have not been followed.

5.1 GENERAL OPERATION

WARNING: Read the Safety information starting on page 11 prior to setting up or operating the GPR-800.

See the component location information in Section 3 for descriptions of GPR-800 components and interfaces.

5.2 OPERATING CONTROLS: KEYPADS AND TIMER RELAYS

The GPR-800 user interface includes two keypads and two timer relays.

> NOTE: Do not use a writing pen or other sharp object when operating the keypad or timer relays as this may damage the component surfaces. Use your finger for all keypad and timer relay selections.

Figure 5-1. Shows the GPR-800 user interface.

5.2.1 Keypads

Each keypad has three control buttons: START, PAUSE, and STOP | SET. Each set of buttons controls 1 bank (CH 1-4 or CH 5-8) of 4 electrodes.

- **START button.**

If no safety interlocks are activated (i.e., the Safety Lid is closed and the Chip Holder is in place), pressing the START button begins the timer countdown and applies a voltage to the corresponding electroelution channels. Additionally, the corresponding Channel LEDs will illuminate, indicating that current is flowing through the channel.

If safety interlocks are activated, pressing the START button transfers the corresponding set of electrodes into a PAUSED state. All channels are identified as paused by blinking Channel LEDs. When the safety interlocks are disengaged (e.g., the safety lid is closed), the system begins the timer countdown and applies a voltage to the corresponding electroelution channels.

- **PAUSE button.**

If the system is delivering electroelution voltage (as indicated by illuminated Channel LEDs), pressing the PAUSE button will pause the timer countdown and remove the applied voltage from the corresponding electroelution channels. Channels which are in a paused state are identified by blinking Channel LEDs.

If no safety interlocks are activated (i.e., the Safety Lid is closed and the Chip Holder is in place) and the system is in a paused state, pressing the PAUSE button resumes the timer countdown and resumes application of voltage to the corresponding electroelution channels. Additionally, the corresponding Channel LEDs will illuminate (not blink), indicating that current is flowing through the channel.

If safety interlocks are activated, pressing the PAUSE button pauses the corresponding Channels. When the safety interlocks are disengaged (e.g., the safety lid is closed), the user must press the START button to resume current delivery.

- **STOP | SET button.**

If the system is delivering electroelution voltage (as indicated by illuminated Channel LEDs), pressing the STOP | SET button will stop the timer countdown, reset the timer value to the initial value, and remove the applied voltage from the corresponding electroelution channels. Channels which are in a stopped state are identified by unlit Channel LEDs.

If the system is paused (as indicated by blinking Channel LEDs), pressing the STOP | SET button will reset the timer value to the initial values and remove the applied voltage from the corresponding electroelution channels. Channels which are in a stopped state are identified by unlit Channel LEDs.

If the system is in a stopped state, pressing the STOP | SET button sets the target time on the timer relays as the relay run time. See Section 5.2.2 for information related to the timer relay interface.

5.2.2 Timer Relays

The GPR-800 has two timer relays (see Figure 5-1). Each relay is dedicated to the control of one bank (CH 1-4 or CH 5-8) of 4 electrodes. Each up/down button of the timer relay (see Figure 5-2) controls one digit of the process time (mm:ss). Pressing the up or down arrows changes the corresponding value of the set time. When the appropriate set time is displayed in the timer, pressing the SET key on the keypad sets the process time to equal the set time.

Figure 5-2. Timer Relay

NOTE: The timer relay set time is displayed in orange below the process time. The timer relay process time is displayed in red above the set time. The process time controls the electroelution time of the GPR-800.

NOTE: The yellow buttons to the left of the timer relay should not be pressed at any time. They are not part of the GPR-800 user interface.

5.3 GEL PROTEIN RECOVERY

The gel protein recovery process steps are described below.

5.3.1 Priming the Protein GPRchip for Gel Protein Recovery

1. Place the chip against a dark background and pipette 200 µL of GPR Electroelution Buffer (GPR-020) into Reservoirs A and B for every channel that will be run. Watch as the buffer completely fills the micro channel and ensure that there are no bubbles present. The hydrodynamic flow will begin as soon as the Reservoirs are filled.

> NOTE: Protea recommends that the GPR buffer be placed in a sonicator for 5 minutes prior to use to minimize the risk of bubbles.

2. Add a cored gel plug to Reservoir A for each channel that will be run. See figure 5-3 for designation of the Reservoirs.

> NOTE: Protea recommends the use of its gel spot cutters to produce 2 mm circular gel spots for optimum efficiency in the GPR-800. However, the GPR-800 chips are compatible with gel spots cut out with razor blades and other coring devices, provided that the gel spots are placed down in the bottom of Reservoir A.

> NOTE: Multiple gel spot cores can be loaded into a single Reservoir to increase the amount of protein available for recovery, but a longer electroelution time may be required.

> NOTE: It is very important to ensure that air bubbles are not present in the microfluidic channels, as air bubbles can cause breakdown of the electrical pathway and current fluctuations during electroelution.

3. Ensure that all gel plugs are at the bottom of Reservoir A for each channel containing a gel sample. An entire bank of electrodes (1-4 or 5-8) must be run together. If you have less than 4 gel pieces, the remaining empty Reservoirs from the unused channels must be filled with 200 µL of de-ionized water before starting the run.

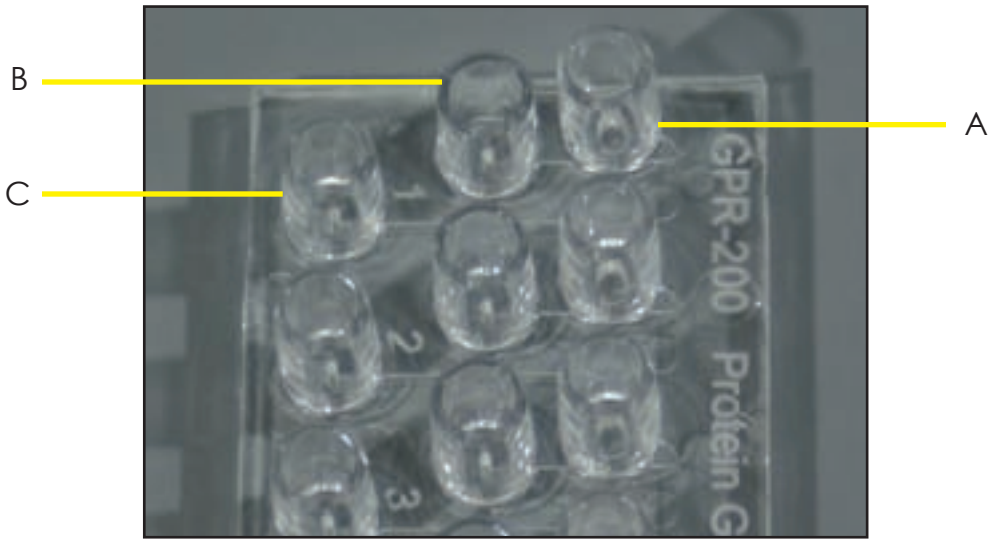

Figure 5-3. Protein GPRchip reservoirs for a single processing channel.

5.3.2 Sample Extraction and Collection

1. Place the microfluidic chip in the GPR-800 chip holder (center chip position, Figure 5-4). Ensure that the chip is pushed down to the bottom left corner in the center position of the holder. Load the holder in its slot inside the instrument.

> NOTE: Protea recommends returning the buffer that has flowed into Reservoir C in equal amounts to Reservoirs A and B. This will minimize the collection volume and maximize the concentration of the recovered protein. If you load the buffer and the gel plugs quickly this will not be necessary

2. Lower the electrode assembly onto the chip and secure the knob in the top hole (resting position) of the metal closure plate. Close the blue safety lid to disengage the safety interlock on the power supply.

> NOTE: The platinum electrodes are fragile and can be bent by rough handling. When working with the electrode assembly or electrodes, please handle gently.

> NOTE: The top latch position is the "resting" (i.e., unsealed) position. The lower latch position seals the reservoirs and should not be used with the Protein GPRchip.

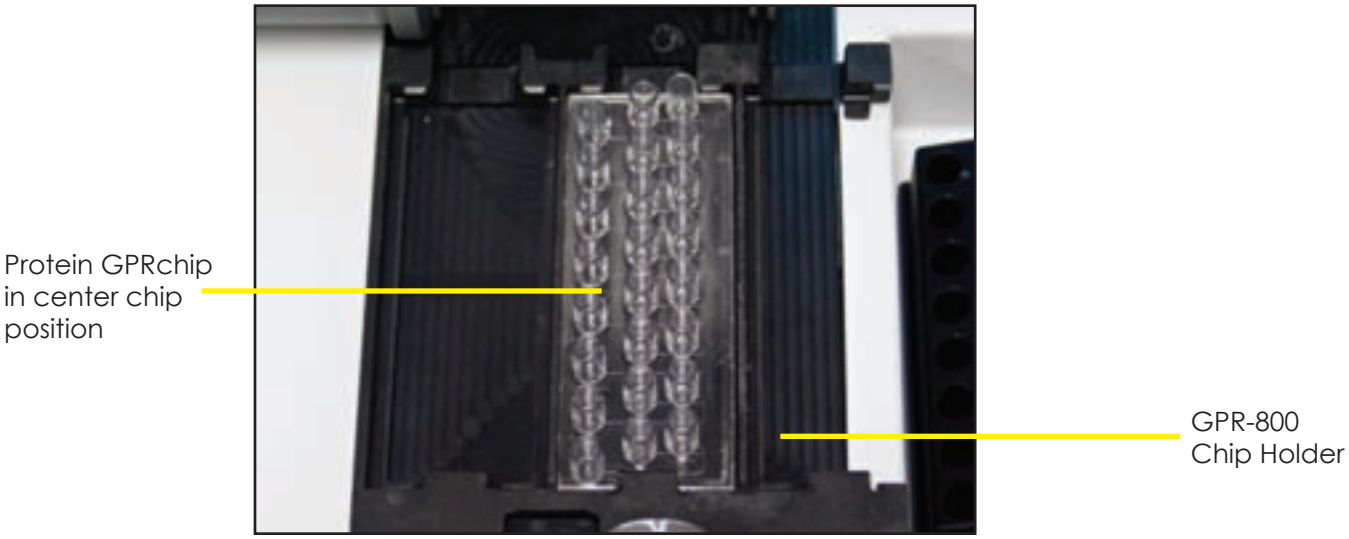

Figure 5-4. Protein GPRchip in the chip holder's center position.

3. Power up the GPR-800 system by turning on the switch at back of instrument. The green POWER STATUS LED will light up to indicate power to the GPR-800.

4. Set the timers to the required electroelution time. Each timer controls a bank of four electrodes (1-4 and 5-8). See Table 5-1 for eletroelution time guidelines.

NOTE: The left timer relay controls and displays the run time for channels 1-4. The right timer relay controls and displays the run time for channels 5-8.

NOTE: The bottom number on the timer relay is the target time, the top number is the run time that will be applied to each bank of channels. When the target time is changed, the STOP | SET button must be pressed to apply this new value as the run time.

Table 5.1. Electroelution Time Guidelines for the GPR-800.

Size of Protein	Time (minutes)	
	Number of Gel Plugs per channel (2 mm, circular)	
	1 Gel Plug	2 Gel Plugs
1-25 kDa	10	20
25-75 kDa	20	30-40
75 kDa and up	20-30	40-60

5. Press the START button(s) to apply the electric field and initiate the gel protein recovery process. When operating the system will apply a constant voltage of 500 V across Reservoirs A and B of each channel. The LED indicators for each channel will change from unlit (OFF) to green when both the high voltage is ON and the current through the channel is above 50 µA.

> NOTE: If the electroelution voltage is ON, the eight channel LEDS will be illuminated. The channel LED will be red if the channel current is <50 µA to indicate the electrical current error. The channel LED will be green if the channel current is >50 µA.

> NOTE: Electroelution can be stopped before the electroelution timer reaches 00:00 by pressing the PAUSE button, pressing the STOP | SET button, or by opening the Safety Cover. Refer to Section 5.3.4 for additional details.

6. When the electroelution time has expired the GPR-800 automatically turns off the electroelution voltage.

7. Sample Collection: Pipette out the sample that collects in Reservoir C into a clean 0.5 mL vial.

5.3.3 Stopping Electroelution

Electroelution will stop when the timer run time reaches 00:00. Electroelution can be stopped before the run time reaches 00:00 by pressing the STOP | SET button at any time.

- If it is necessary to shut off the electroelution voltage during the gel protein recovery process, either push the PAUSE button or open the blue safety lid to automatically place the GPR-800 in a pause mode.

- When the pause feature is engaged, the LED's will blink red. If the PAUSE button is depressed to pause the instrument, then press START to re-start the sample processing at the time point where the pause was initiated.
- If the blue safety lid is opened to pause the instrument, the voltage will be restored when the lid is closed.

5.3.4 LC-MS Top-Down Analysis

1. Sample collection and detergent degradation:

- Collect sample from protein GPRchip into microcentrifuge tube using a micropipette.
- Add Surfactant Degradation Reagent (10X) to the sample to achieve a 10-fold dilution (e.g. add ~15 µL of Surfactant Degradation Reagent to a 150 µL GPR sample) and incubate at room temperature for 15 – 30 minutes (15 minutes for MALDI analysis ; 30 minutes for ESI analysis).

2. Desalt protein sample using a C_8 SpinTip:

- Ensure that the packing material is at the bottom of the tip by gently tapping the tip to displace any packing material sticking to the top cap.
- Place a centrifuge adaptor onto a 2 mL centrifuge tube.
- Remove the cap from the SpinTip and place into the centrifuge adaptor.
- Wash the SpinTip to wet the packing material by adding 50 µL of Equilibration Solution to the top of the SpinTip using a micropipette. Centrifuge the system at 4000 x g for 3 min. Repeat the SpinTip wash.
- Rinse the SpinTip by adding 50 µL of Sample Reconstitution and Rinse Solution to the top of the SpinTip. Centrifuge the system at 4000 x g for 3 min. Repeat the SpinTip rinse.
- Load 10 to 200 µL of the GPR Sample Solution by adding it to the top of the SpinTip and centrifuging the system at 4000 x g for 3 min. Additional sample volumes can be added in 200 µL aliquot cycles.
- Wash the sample to elute salts and other non-retained components by adding 100 µL of the GPR Rinse Buffer for ESI to the top of the SpinTip. Centrifuge the system at 4000 x g for 3 min. Repeat the SpinTip sample wash.
- Transfer the SpinTip and adapter to a new clean centrifuge tube to collect the sample during elution.

- Elute the sample by adding 100 µL of Elution Solution to the top of the SpinTip. Centrifuge the system at 4000 x for 3 min.

  NOTE: Monitor the level of liquid in the waste centrifuge tube being careful the flow through volume does not cover the bottom point of the SpinTip.

- Dry down sample in a lyophilizer or SpeedVac.

  NOTE: for proteins > 75 kDa, a C_4 SpinTip should be used

3. LC-MS Analysis of Intact Proteins

- Add 10-20 µL of the LC aqueous (sample loading) solution to the solid sample and sonicate briefly to dissolve the sample.
- Load sample into sample loop on LC-MS system.
- Load sample from sample loop onto C_8 column and begin LC-MS analysis.

5.3.5 MALDI Intact Mass Measurement Prep Analysis

1. Sample collection and detergent degradation:

- Collect sample from protein GPRchip into microcentrifuge tube using a micropipette.
- Add Surfactant Degradation Reagent (10X) to the sample to achieve a 10-fold dilution (e.g. add ~15 µL of Surfactant Degradation Reagent to a 150 µL GPR sample) and incubate at room temperature for 15 – 30 minutes (15 minutes for MALDI analysis; 30 minutes for ESI analysis).

2. Desalt protein sample using a C_8 SpinTip:

- Ensure that the packing material is at the bottom of the tip by gently tapping the tip to displace any packing material sticking to the top cap.
- Place a centrifuge adaptor onto a 2 mL centrifuge tube.
- Remove the cap from the SpinTip and place into the centrifuge adaptor.
- Wash the SpinTip to wet the packing material by adding 50 µL of Equilibration Solution to the top of the SpinTip using a micropipette. Centrifuge the system at 4000 x g for 3 min. Repeat the SpinTip wash.
- Rinse the SpinTip by adding 50 µL of Sample Reconstitution and Rinse Solution to the top of the SpinTip. Centrifuge the system at 4000 x g for 3 min. Repeat the SpinTip rinse.
- Load 10 to 200 µL of the GPR Sample Solution by adding it to the top of the SpinTip and centrifuging the system at 4000 x g for 3 min. Additional sample volumes can be added in 200 µL aliquot cycles.

- Wash the sample to elute salts and other non-retained components by adding 100 µL of the GPR Rinse Buffer for MALDI to the top of the SpinTip. Centrifuge the system at 4000 x g for 3 min. Repeat the SpinTip sample wash.
- Transfer the SpinTip and adapter to a new clean centrifuge tube to collect the sample during elution.
- Elute the sample by adding 100 µL of Elution Solution to the top of the SpinTip. Centrifuge the system at 4000 x g for 3 min.

  NOTE: Monitor the level of liquid in the waste centrifuge tube being careful the flow through volume does not cover the bottom point of the SpinTip.

- Dry down sample in a lyophilizer or SpeedVac.

  NOTE: for proteins > 75 kDa, a C_4 SpinTip should be used

3) MALDI Analysis of Intact Proteins

- Add 10-20 µL of the Sample Reconstitution and Rinse Solution to the solid sample and sonicate briefly to dissolve the sample.
- Wash and equilibrate a C_8 LithTip using the Equilibration Buffer. Add 25 µL of buffer to a clean vial and then perform 10 cycles of aspiration and expulsion of the buffer using the micropipette set at 10 µL.
- Rinse the C_8 LithTip using Reconstitution and Rinse Solution. Add 25 µL of buffer to a clean vial and then perform 10 cycles of aspiration and expulsion of the buffer using the micropipette set at 10 µL.
- Load the sample into the C8 LithTip by aspirating and expelling the sample 5-10 times within the sample vial.
- Wash the sample on the C_8 LithTip with GPR Rinse Buffer for MALDI.
- Add 25 µL of buffer to a clean vial and then perform 10 cycles of aspiration of the buffer using the micropipette set at 10 µL.
- Elute the sample with 2 µL of Elution Solution, by spotting the 2 µL directly onto the MALDI target, pre-spotted with 0.5 µL MALDI matrix (e.g. CHCA) and allowing to air-dry.
- Load the MALDI target into the mass spectrometer and begin mass spec analysis.

5.3.6 Bottom-Up Analysis by ESI (Post-GPR Digestion)

1. Sample collection and detergent degradation:

- Collect sample from protein GPRchip into microcentrifuge tube using a micropipette.
- Add Surfactant Degradation Reagent (10X) to the sample to achieve a 10-fold dilution (e.g. add ~15 µL of Surfactant Degradation Reagent to a 150 µL GPR sample) and incubate at room temperature for 15 – 30 minutes (15 minutes for MALDI analysis ; 30 minutes for ESI analysis).

2. Desalt protein sample using a C_8 SpinTip:

- Ensure that the packing material is at the bottom of the tip by gently tapping the tip to displace any packing material sticking to the top cap.
- Place a centrifuge adaptor onto a 2 mL centrifuge tube.
- Remove the cap from the SpinTip and place into the centrifuge adaptor.
- Wash the SpinTip to wet the packing material by adding 50 µL of Equilibration Solution to the top of the SpinTip using a micropipette. Centrifuge the system at 4000 x g for 3 min. Repeat the SpinTip wash.
- Rinse the SpinTip by adding 50 µL of Sample Reconstitution and Rinse Solution to the top of the SpinTip. Centrifuge the system at 4000 x g for 3 min. Repeat the SpinTip rinse.
- Load 10 to 200 µL of the GPR Sample Solution by adding it to the top of the SpinTip and centrifuging the system at 4000 x g for 3 min. Additional sample volumes can be added in 200 µL aliquot cycles.
- Wash the sample to elute salts and other non-retained components by adding 100 µL of the GPR Rinse Buffer for ESI to the top of the SpinTip. Centrifuge the system at 4000 x g for 3 min. Repeat the SpinTip sample wash.
- Transfer the SpinTip and adapter to a new clean centrifuge tube to collect the sample during elution.
- Elute the sample by adding 100 µL of Elution Solution to the top of the SpinTip. Centrifuge the system at 4000 x g for 3 min.

  NOTE: If the sample is targeted for LC-MS analysis by electrospray ionization (ESI) mass spectrometry, then the Elution Solution for ESI should be used (formic acid based).

  NOTE: Monitor the level of liquid in the waste centrifuge tube being careful the flow through volume does not cover the bottom point of the SpinTip.

- Dry down sample in a lyophilizer or SpeedVac.

  NOTE: for proteins > 75 kDa, a C_4 SpinTip should be used

3. Solution-phase digestion:

- Dissolve protein pellet in 100 µL of Digestion Buffer (50 mM ammonium bicarbonate in 20% acetonitrile).
- Vortex or sonicate the tube to aid in solubilization. Using a centrifuge, spin the tube at 5,000g for 1 minute to make sure that the sample solution is collected in the bottom of the tube.
- Reduction → DTT:
- Add 5 µL of 250 mM DTT (in 50 mM ammonium bicarbonate in 20% acetonitrile) to the sample solution. This dilution will give a final concentration of 10 mM DTT.

  NOTE: Add 0.386 g DTT to 10 mL of 50 mM ammonium bicarbonate in 20% acetonitrile to make the 250 mM solution.

- Incubate at 40˚C for 1 hour on heat plate.
- Alkylation → Iodoacetamide:
- Add 5 µL of 625 mM iodoacetamide (in 50 mM ammonium bicarbonate in 20% acetonitrile) to the sample solution. This dilution results in a final concentration of 25 mM iodoacetamide.

  NOTE: Add 1.156 g iodoacetamide to 10 mL of 50 mM ammonium bicarbonate in 20% acetonitrile to make the 625 mM solution.

- Incubate at room temperature in the dark for 1 hour.
- Add 2.0 µL of trypsin solution (1.0 µg/µL stock solution) to the reduced and alkylated sample solution.
- Incubate overnight (or for 12 hours) at 37˚C on heat plate.
- Stop the digestion by adding 10 µL of glacial acetic acid.
- The digestion solution can be transferred to a labeled 1.5 mL centrifuge tube and dried down in a lyophilizer or SpeedVac to produce the crystallized peptides.
- Freeze the peptides at -80˚C until ready for use.
- For MALDI analysis, the protein digest can be spotted directly on the plate or with a C_{18} LithTip using CHCA matrix
- For LC-MS or LC-MALDI analysis, the protein digest can be reconstituted in LC aqueous buffer and then loaded onto the chromatographic column for separation and analysis.

5.3.7 Bottom-Up Analysis by MALDI (Post-GPR Digestion)

1. Sample collection and detergent degradation:

- Collect sample from protein GPRchip into microcentrifuge tube using a micropipette.
- Add Surfactant Degradation Reagent (10X) to the sample to achieve a 10-fold dilution (e.g. add ~15 µL of Surfactant Degradation Reagent to a 150 µL GPR sample) and incubate at room temperature for 15 – 30 minutes (15 minutes for MALDI analysis; 30 minutes for ESI analysis).

2. Desalt protein sample using a C_8 SpinTip:

- Ensure that the packing material is at the bottom of the tip by gently tapping the tip to displace any packing material sticking to the top cap.
- Place a centrifuge adaptor onto a 2 mL centrifuge tube.
- Remove the cap from the SpinTip and place into the centrifuge adaptor.
- Wash the SpinTip to wet the packing material by adding 50 µL of Equilibration Solution to the top of the SpinTip using a micropipette. Centrifuge the system at 4000 x g for 3 min. Repeat the SpinTip wash.
- Rinse the SpinTip by adding 50 µL of Sample Reconstitution and Rinse Solution to the top of the SpinTip. Centrifuge the system at 4000 x g for 3 min. Repeat the SpinTip rinse.
- Load 10 to 200 µL of the GPR Sample Solution by adding it to the top of the SpinTip and centrifuging the system at 4000 x g for 3 min. Additional sample volumes can be added in 200 µL aliquot cycles.
- Wash the sample to elute salts and other non-retained components by adding 100 µL of the GPR Rinse Buffer for MALDI to the top of the SpinTip. Centrifuge the system at 4000 x g for 3 min. Repeat the SpinTip sample wash.
- Transfer the SpinTip and adapter to a new clean centrifuge tube to collect the sample during elution.
- Elute the sample by adding 100 µL of Elution Solution to the top of the SpinTip. Centrifuge the system at 4000 x g for 3 min.

  NOTE: If the sample is targeted for LC-MS analysis by electrospray ionization (ESI) mass spectrometry, then the Elution Solution for ESI should be used (formic acid based).

  NOTE: Monitor the level of liquid in the waste centrifuge tube being careful the flow through volume does not cover the bottom point of the SpinTip.

- Dry down sample in a lyophilizer or SpeedVac.

  NOTE: for proteins > 75 kDa, a C_4 SpinTip should be used

3. Solution-phase digestion:

- Dissolve protein pellet in 100 µL of Digestion Buffer (50 mM ammonium bicarbonate in 20% acetonitrile).
- Vortex or sonicate the tube to aid in solubilization. Using a centrifuge, spin the tube at 5,000g for 1 minute to make sure that the sample solution is collected in the bottom of the tube.
- Reduction → DTT:
- Add 5 µL of 250 mM DTT (in 50 mM ammonium bicarbonate in 20% acetonitrile) to the sample solution. This dilution will give a final concentration of 10 mM DTT.

  NOTE: Add 0.386 g DTT to 10 mL of 50 mM ammonium bicarbonate in 20% acetonitrile to make the 250 mM solution.

- Incubate at 40˚C for 1 hour on heat plate.
- Alkylation → Iodoacetamide:
- Add 5 µL of 625 mM iodoacetamide (in 50 mM ammonium bicarbonate in 20% acetonitrile) to the sample solution. This dilution results in a final concentration of 25 mM iodoacetamide.

  NOTE: Add 1.156 g iodoacetamide to 10 mL of 50 mM ammonium bicarbonate in 20% acetonitrile to make the 625 mM solution.

- Incubate at room temperature in the dark for 1 hour.
- Add 2.0 µL of trypsin solution (1.0 µg/µL stock solution) to the reduced and alkylated sample solution.
- Incubate overnight (or for 12 hours) at 37˚C on heat plate.
- Stop the digestion by adding 10 µL of glacial acetic acid.
- The digestion solution can be transferred to a labeled 1.5 mL centrifuge tube and dried down in a lyophilizer or SpeedVac to produce the crystallized peptides.
- Freeze the peptides at -80˚C until ready for use.
- For MALDI analysis, the protein digest can be spotted directly on the plate or with a C_{18} LithTip using CHCA matrix
- For LC-MS or LC-MALDI analysis, the protein digest can be reconstituted in LC aqueous buffer and then loaded onto the chromatographic column for separation and analysis.

5.4 POWER OFF

At any time the user may remove power from the GPR-800 by toggling the main power switch on the rear of the unit.

NOTE: Toggling the main power switch on the rear of the unit removes all power from the GPR-800.

NOTE: The timer relay set time (displayed in orange below the process time) at the time the unit is powered off will become the process time when the unit is powered on. See Section 5.2.2 for further descriptions of the timer relay set and process times.

6 Cleaning and Maintenance

6.1 CLEANING

After each use of the GPR-800, clean the electrodes, electrode hinge, vial lift assembly, chip holder, vial holder, and exterior surfaces of the GPR-800. Surfaces should be cleaned with a lint-free wipe dampened with isopropyl alcohol. Use care not to wet any internal electronic components or exposed wiring. Also take care not to bend the electrodes.

WARNING: Turn off electrical power to the system and unplug the power supply cord before cleaning the GPR-800.

CAUTION: The system electrodes are delicate and sharp. Use care when working around the electrodes.

6.2 MAINTENANCE

No aspect of the GPR-800 is serviceable by the end user. Only a Protea trained service technician should perform maintenance on the GPR-800.

CAUTION: Users should not attempt to repair the GPR-800. The GPR-800 is designed to be serviceable only by Protea trained service technicians. (877.776.8321).

Table 7-1. Troubleshooting Guide

Problem	Cause	Resolution
Blinking red LEDs	System is paused Blue safety lid is open Chip holder not properly placed	Press start to begin the run Lower lid, run will start automatically Make sure the chip holder is in it proper place
One or more channels shows red LED at the start of the run	Insufficient current is flowing through that channel due to a bubble in the micro-channel Insufficient volume in the buffer reservoir	Aspirate the buffer from the collection reservoir to remove the bubble. Add the aspirated buffer back to the electrode reservoirs Check to ensure buffer was placed in all 4 or 8 channels. Top-up buffer in buffer reservoirs
One or more channels shows a red LED during the run	A bubble has formed or entered the micro-channel during the run	In most cases the bubble will move through the channel and the current will return to normal. If the LED is red for a prolonged period of time (several minutes), re-run that channel
One or more channels LEDs do not light up at the start of the run	Blown circuit board fuse	Contact Protea technical service for guidance in replacing of circuit board fuse
Coomassie stained gels still colored at the end of the run	Incomplete electroelution Electroelution from an old gel with proteins that are strongly fixed in the gel	Increase electroelution time Electroelute proteins run on fresh gels
Difficulty in recovering high molecular weight proteins	Slow movement of proteins out of the polyacrylamide gel	Electroelute proteins run on a lower percent acrylamide gel

Table 7-2. Error Message Guide

Error Message	Cause	Solution
Device Not Found	Communication with the GPR-800 data acquisition device has been lost	Ensure that the GPR-800 is powered on (Power Status LED is lit) and that the USB cable is properly connected to the GPR-800 and the computer running the GPR-800 software.
USB connection is not responding…	Communication with the GPR-800 data acquisition device has been lost.	See resolution to "Device Not Found" error / warning.
Calibration file not found.	Device-specific calibration file has been deleted, relocated, or renamed.	I Input device-specific calibration values when prompted by the software.

GPR-800 Reagent Bundles and Kits

GPR LC-MS Top-Down Prep Kit
Contains Surfactant Degradation Reagent (25 mL) and C_8 GPR SpinTips Sample Prep Kit for ESI (24 pack)

Catalog Number	Size	Price
GPR-050	ea.	$145

GPR MALDI Intact Mass Prep Kit
Contains Surfactant Degradation Reagent (25 mL), C_8 GPR SpinTip Sample Prep Kit for MALDI (24 pack), C_8 LithTip Sample Prep Kit (24 pack), and Ultrapure CHCA MALDI Matrix (5 x 10mg vials)

Catalog Number	Size	Price
GPR-055	ea.	$275

GPR LC-MS Bottom-Up Prep Kit
Contains Surfactant Degradation Reagent (25 mL) and C_8 GPR SpinTips Sample Prep Kit for ESI, and Trypsin Digestion Kit

Catalog Number	Size	Price
GPR-060	ea.	$245

GPR MALDI Bottom-Up Prep Kit
Contains Surfactant Degradation Reagent (25 mL), C_8 GPR SpinTip Kit for MALDI (24 pack), C_8 LithTip Sample Prep Kit (24 pack), Ultrapure CHCA MALDI Matrix (5 x 10mg vials), and Trypsin Digestion Kit

Catalog Number	Size	Price
GPR-065	ea.	$375

GPR-800 Reagent Bundle, Protein Recovery
Contains Protein GPRchips (10)and GPR Electroelution buffer (2 x 25 mL)

Catalog Number	Size	Price
GPR-070	ea.	$550

GPR-800 Accessory Kit
Contains spot cutter holder, metal spot cutter cores with 0-rings (50), and forceps (1 pair)

Catalog Number	Size	Price
GPR-075	ea.	$145

GPR-800 Accessories

Protein GPRchip
Contains 8 channels for protein electroelution from polyacrylamide gel samples, PMMA, pack of 10

Catalog Number	Size	Price
GPR-200-10	10-pk	$450

Protein GPRchip
Contains 8 channels for protein electroelution from polyacrylamide gel samples, PMMA, pack of 25

Catalog Number	Size	Price
GPR-200-25	25-pk	$1000

GPR Electroelution Buffer

Catalog Number	Size	Price
GPR-020-25mL	25 mL	$95
GPR-020-4x25mL	4X25 mL	$295

Surfactant Degradation Reagent for MALDI, 10 mL

Catalog Number	Size	Price
GPR-025-10mL	10 mL	$45

Surfactant Degradation Reagent for ESI, 10 mL

Catalog Number	Size	Price
GPR-026-10mL	10 mL	$45

Spot Cutter Holder

Catalog Number	Size	Price
GPR-030	ea.	$50

Metal spot cutter cores with o-rings, 50 pk

Catalog Number	Size	Price
GPR-035-50	pk.	$100

GPR-800 Accessories

Forceps, 1 pair

Catalog Number	Size	Price
GPR-040	ea.	$12

C_8 LithTips Sample Prep Kit, 1-10 µL, 24 Tip Kit

Kit contains C_8 LithTips, Sample Reconstitution and Rinse Solution, Equilibration Solution, and Elution Solution

Catalog Number	Size	Price
SP-420-24	24 tip kit	$95

C_8 LithTips Sample Prep Kit, 1-10 µL, 96 Tip Kit

Kit contains C_8 LithTips, Sample Reconstitution and Rinse Solution, Equilibration Solution, and Elution Solution

Catalog Number	Size	Price
SP-420-96	96 tip kit	$345

C_8 GPR SpinTips Sample Prep Kit for ESI, 24 Tip Kit

Kit contains C_8 LithTips, Sample Reconstitution and Rinse Solution, GPR Rinse Buffer for ESI, Equilibration Solution, and Elution Solution

Catalog Number	Size	Price
SP-161-24	24 tip kit	$125

C_8 GPR SpinTips Sample Prep Kit for ESI, 24 Tip Kit

Kit contains C_8 LithTips, Sample Reconstitution and Rinse Solution, GPR Rinse Buffer for ESI, Equilibration Solution, and Elution Solution

Catalog Number	Size	Price
SP-161-96	96 tip kit	$375

C_8 GPR SpinTips Sample Prep Kit for MALDI, 24 Tip Kit

Kit contains C_8 LithTips, Sample Reconstitution and Rinse Solution, GPR Rinse Buffer for MALDI, Equilibration Solution, and Elution Solution

Catalog Number	Size	Price
SP-171-24	24 tip kit	$125

C_8 GPR SpinTips Sample Prep Kit for MALDI, 96 Tip Kit

Kit contains C_8 LithTips, Sample Reconstitution and Rinse Solution, GPR Rinse Buffer for MALDI, Equilibration Solution, and Elution Solution

Catalog Number	Size	Price
SP-171-96	96 tip kit	$375

C_4 LithTips Sample Prep Kit, 1-10 µL, 24 Tip Kit

Kit contains C_4 LithTips, Sample Reconstitution and Rinse Solution, Equilibration Solution, and Elution Solution

Catalog Number	Size	Price
SP-410-24	24 tip kit	$95

C_4 LithTips Sample Prep Kit, 1-10 µL, 96 Tip Kit

Kit contains C_4 LithTips, Sample Reconstitution and Rinse Solution, Equilibration Solution, and Elution Solution

Catalog Number	Size	Price
SP-410-96	96 tip kit	$345

C_4 GPR SpinTips Sample Prep Kit for ESI, 24 Tip Kit

Kit contains C_4 LithTips, Sample Reconstitution and Rinse Solution, GPR Rinse Buffer for ESI, Equilibration Solution, and Elution Solution

Catalog Number	Size	Price
SP-162-24	24 tip kit	$125

C_4 GPR SpinTips Sample Prep Kit for ESI, 24 Tip Kit

Kit contains C_4 LithTips, Sample Reconstitution and Rinse Solution, GPR Rinse Buffer for ESI, Equilibration Solution, and Elution Solution

Catalog Number	Size	Price
SP-162-96	96 tip kit	$375

C_4 GPR SpinTips Sample Prep Kit for MALDI, 24 Tip Kit

Kit contains C_4 LithTips, Sample Reconstitution and Rinse Solution, GPR Rinse Buffer for MALDI, Equilibration Solution, and Elution Solution

Catalog Number	Size	Price
SP-172-24	24 tip kit	$125

C_4 GPR SpinTips Sample Prep Kit for MALDI, 96 Tip Kit

Kit contains C_4 LithTips, Sample Reconstitution and Rinse Solution, GPR Rinse Buffer for MALDI, Equilibration Solution, and Elution Solution

Catalog Number	Size	Price
SP-172-96	96 tip kit	$375

9.1 SDS-PAGE Gel Recommendations for GPR-800 Samples

I. SDS-PAGE gel
Recommended gel vendors

- Protea Biosciences (ProteaGels)
- BioRad (Mini-Protean and Criterion gels)
- Life Technologies, formerly Invitrogen (Novex NuPAGE gels)

Supported gel characteristics
Percent acrylamide

- 5, 10, 12, and 15% polyacrylamide
- 10 and 12% gels work best for most proteins
- 15% gels should not be used for proteins over 30 kDa

Crosslinker ratio

- Higher polyacrylamide monomer:crosslinker ratios produce larger average pore sizes that improve gel protein recovery
- Recovery trends: 37.5:1 > 29:1 > 19:1
- Non-supported gel characteristics
- Gradient gels – gradient gels have variable pore sizes across the length of the gels. This variation in pore size can have an unpredictable effect on the recovery of proteins by the GPR-800.
- While some proteins may be not affected or even positively influenced by the use of a gradient gel with the GPR-800, many proteins can be located in gel regions containing greater than 12% acrylamide, which increases the resistance of electrophoretic movement within the gel and decreases the efficiency of recovery.
- Variations in the gel vendor, type of gradient (linear vs non-linear), percent gradient (e.g. 5 – 20%), lot-to-lot reproducibility, and length of electrophoretic run can all significantly influence the reproducibility and efficiency of gel protein recovery for gradient gel samples.
- Gradient gels should only be used for advanced GPR-800 users who are performing optimized methods development for gel protein recovery. Consequently, Protea Biosciences does not support the use of gradient gels for routine gel protein recovery.

2. Electrophoresis

- SDS-PAGE gels should be run according to the manufacturer's instructions for voltages, currents, and run times.
- In general, run voltages of 100 to 180V are recommended.
- For run-to-run reproducibility in both the protein gel separation and gel protein recovery, the SDS-PAGE gels should be run until the dye front reaches the bottom of the gel.

NOTES: If the run time is too short for complex samples, then the sample proteins may not fully resolve, leading to protein bands that may contain multiple protein species – all of which may be recovered during gel protein recovery.

NOTES: This recommendation is very important for investigators who opt to utilize gradient gels, as the pore size of the gel varies along the length of the gel as the percent acrylamide changes.

3. Staining and visualization

Supported stains

- Zinc
- Sypro Ruby
- Coomassie
  - R-250 (non-colloidal)
  - G-250 (colloidal)

Non-supported stains

- Silver
- Imperial blue
- Colloidal gold

Gel Fixation

- Fixation of proteins after SDS-PAGE separation is often performed to prevent diffusional protein movement with the gel (loss of resolution) and migration out of the gel (protein sample loss from the gel).
- Fixation of the proteins is a precipitation event that is typically induced by using a fixing solution containing high percentages or organic solvent and/or organic acids.

Supported fixing solutions:

- 10% Methanol 7% Acetic Acid (SYPRO Ruby protein gel stain)
- 50% Methanol 10% Acetic Acid (Coomassie Blue R250 stain)
- 0.1% Methanol 5% Phosphoric Acid (Coomassie Blue G250 stain)

NOTE: The staining methods in parentheses above are the methods that typically call for these individual fixing solution recipes. Each fixing solution can be used for other staining techniques.

- Harsher protein fixation protocols can produce non-reversible protein precipitations that lead to reduced efficiency and/or inhibition of both in-gel digestion and gel protein recovery.

NOTE: The use of acetonitrile is not recommended for protein fixation as it is very effective at precipitating proteins >50 kDa at concentrations >50%.

Gel Destaining

- Zinc and SYPRO Ruby stained protein gels do not require destaining and can be used directly for gel protein recovery.
- Coomassie stained protein gels that are used for gel protein recovery will recover the Coomassie blue dye along with the protein. This dye will produce an intense chromatographic peak, and it may also lead to the production of Coomassie adducts in the mass spec.

NOTE: At this time, there is not a recommended destain protocol for use with Coomassie stained protein gels.

NOTE: The use of acetonitrile-based destain protocols can lead to the precipitation of the proteins in the gel matrix and/or non-reproducible gel protein recovery.

Protocols

Zinc stain

- Remove the gel from the cassette once gel electrophoresis is completed and place in an appropriate size staining container.
- Add 50 mL of 0.2 M imidazole solution.

  NOTE: Use sufficient volume to immerse gel

- Equilibrate on shaker for 10 minutes.
- Discard imidazole solution.
- Add 50 mL 0.2 M zinc sulfate solution and shake tray rapidly for 10-60 seconds. The gel becomes opaque white and the protein spots remain transparent.

  NOTE: The color transition time is sample dependent. Larger gel protein loads can develop the gel spots in as little as 10 seconds. Smaller gel protein loads may take 60 seconds or longer.

  NOTE: If the gel is incubated for too long in the zinc sulfate solution, it will over-develop and the protein spots will develop the same opaque white color as the gel background.

  NOTE: Place the receptacle over a dark background to observe the staining reaction and stop it at the correct time point before over-development.

- Rinse the gel in 100 mL ddH_2O for 3 minutes.
- Replace with fresh ddH_2O for storage.

Sypro Ruby fluorescent stain

- Remove the gel from the cassette once gel electrophoresis is completed and place in an appropriate size staining container.
- Wash gel with deionized water to remove residual TGS running buffer.
- Add 50 mL deionized water and shake gently for 30 seconds
- Discard rinse water.
- Add 50 mL of SYPRO Ruby protein gel stain to cover the gel.

  NOTE: Use sufficient SYPRO Ruby protein gel stain to sufficiently cover the gel. In general, use ten times the volume of the gel. Using too little stain will reduce sensitivity.

- Stain the gel with continuous gentle agitation for at least 3 hours for maximal sensitivity.

  NOTE: Specific staining can be seen within 30-90 minutes.

  NOTE: For convenience, gels may be left in the stain overnight (16-18 hours) without overstaining.

- Rinse the gel in 50-100 mL Rinse Solution (10% methanol, 7% acetic acid) for 30-60 minutes.

  NOTE: This rinse step decreases background fluorescence.

  NOTE: Alternatively, 10% ethanol can be used to replace the 10% methanol.

- Wash gel with deionized water prior to imaging.
- Add 50 mL deionized water and shake gently for 30 seconds
- Discard rinse water.
- Add fresh 50 mL deionized water.

Gel imaging

- SYPRO Ruby protein gel stain has two excitation peaks at ~280 nm and ~450 nm and has an emission maxima near 610 nm.
- Stained proteins can be visualized using a variety of excitation sources, including a 300 nm UV or blue-light transilluminator, or laser-based systems.
- SYPRO Ruby protein gel stain also has exceptional photostability and a long emission lifetime, allowing for long exposure times while minimizing background fluorescence.

Coomassie R-250 stain (SB-R250)

- Remove the gel from the cassette once gel electrophoresis is completed and place in an appropriate size staining container containing 50 mL 1X Coomassie R-250 solution.
- Gently shake for 45-60 minutes.
- Remove the Coomassie solution from the gel and dispose in an appropriate waste container.
- Add 50 mL of fixation solution to the gel and continue shaking for an additional 30 minutes.
- Dilute the fixation solution by adding ~100 mL of ddH_2O and add a folded paper towel to absorb additional Coomassie from the solution.
- Gently shake until the required intensity of protein staining is accomplished and background staining is reduced. This step will take several hours.

  NOTE: When using ProteaGels do not leave gels on final step overnight because it will de-stain your protein bands

Reagent recipes:

- 1X Coomassie R-250
- 400 mL ddH_2O
- 500 mL methanol
- 100 mL acetic acid
- 110 mL of 10X Coomassie R-250 stock solution

Fixation solution

- 400 mL ddH_2O
- 500 mL methanol
- 100 mL acetic acid

Coomassie G-250 stain (SB-G250)

- Remove the gel from the cassette once gel electrophoresis is completed and place in an appropriate size staining container containing 100 ml ddH_2O.
- Gently shake for 5 minutes.
- Replace with fresh ddH_2O and wash an additional 2 times for 5 minutes each.
- Remove the ddH_2O from the gel and discard.
- Add 50 mL Coomassie G-250 to the gel and continue shaking for an additional 45-60 minutes.
- Remove the Coomassie solution from the gel and dispose in an appropriate waste container.
- Add 50 mL ddH_2O to the gel and continue shaking for additional 30 minutes.
- Gently shake until the required intensity of protein staining is accomplished and background staining is reduced. May be left overnight.

4. Gel sample handling and storage

- Gel protein recovery experiments produce the highest recoveries for freshly run gels. When possible, it is always recommended that the SDS-PAGE gel separation and gel protein recovery be performed on the same day.
- Gel storage: It is recommend to store whole gels at 4 °C in a minimal amount to water to reduce diffusional protein losses.

  NOTE: Do not freeze the gels, as the formation of ice crystals within and around the gel can damage the gel matrix, leading to sample loss and reduced efficiency of gel protein recovery.

- Over time, proteins will diffuse out of gel stored at 4 °C and leach into the water. It is recommended to use gels for gel protein recovery that have been stored properly for less than two weeks. Gels older than two weeks are not recommended for typical gel protein recovery experiments.